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Long non-coding RNAs in cellular networks and genome regulation

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Trieste, Italy ICGEB (AREA science Park)



Starting with my passion to DNA sequencing in late 80's

Lesson learned:

Technologies make revolutions

... Often leading far from hypotheses

Technology development and the establishment of FANTOM

- @ Yoshihide Hayashizaki's lab at RIKEN (1995)
 - RIKEN wanted to develop technology to identify genes in genome-wide scale.
- How to construct a catalogue of genes;
 How genes are regulated
 - (1) Full-length cDNA technologies
 - (2) Mapping regulatory elements: Cap Analysis of Gene Expression (CAGE; later in the presentation)



Where to go, as RIKEN/Japan?



How to analyze lots of sequences??

We made the project (the experimental part at least)!

Libraries from all mouse tissues/developmental stages,

Millions cDNAs (= expressed genes) end-sequenced

- >103,000 cDNAs fully sequenced
 How to analyze so much data?
 - Call your friends
 - Make new friends!
 - Bold proposal: invite many new friends to look at the data
- FANTOM Project:

Functional ANnoTation Of Mammalian Genome





Figure 1 A shot from the Zen meditation ceremony held as an excursion during the FANTOM2 Cherry Blossom Meeting. The Zen meditation was a good break and provided the participants with novel inspiration.

Unexpected discoveries

There is much more than what in textbooks

Expectations ←???→ Data



New reality- many more RNAs (mRNA, other "non coding" RNAs)

Discovery of "non-coding" RNA Still we know very little



Charting genome output: gene forests, IncRNAs, antisense



Regions of the genome are used multiple times, multiple RNAs as output

The new transcriptional landscape



The RNA world is crowdy: "DARK MATTER" of the transcriptome?



"transcriptome" refers to the collection of all the RNA transcripts

Controversy? from 2002- 2012 and beyond

✓ Some attacks

- ✓ The RIKEN cDNA libraries are full of "junk".
 → Discovery of IncRNAs
- ✓ The RIKEN library are full of cDNA in the wrong orientation.
 → Discovery of antisense RNAs.
- \checkmark Papers attacking the findings
- \checkmark Papers supporting the findings
- ✓ Subsequent papers confirming the findings

"Dogma" versus evidence from new data

The genetic basis of developmental complexity



- Humans (and other vertebrates) have approximately the same number of protein-coding genes (~20,000) as *C. elegans.*
- Most of the proteins are orthologous and have similar functions from nematodes to humans, and many are common with yeast.
- Where is the information that programs our complexity?

Slide courtesy of John Mattick

Next to the "gene catalogue": How to broadly study gene regulation?

- Long journey to map regulatory elements and identify, characterize and use novel RNAs
- Technologies $\leftarrow \rightarrow$ biology and medicine

Understanding regulatory elements in genome



CAGE to broadly map promoters (and enhancers) \rightarrow 5' UTRs

CAGE

Cap Analysis of Gene Expression



- Precise identification of TSSs.
- Quantitative analysis of TSS activity and promoter maps
- Genome-wide

FANTOM5: Regulatory elements in most primary cell types



Forrest et al. Nature 507, 462 (2014)

Overview of approaches; 3000 libraries



Forrest et al. Nature 507, 462 (2014)

Expected/unexpected results

Measure gene expression
 Infer networks globally
 Map promoters



FANTOM5: *Promoter architectures differ in different cells*



B4GALT1 Astrocyte donor1 Astrocyte donor2 Astrocyte donor3 TT CD14+ donor1 CD14+ donor2 TT CD14+ donor2 TT CD14+ donor1 CD14+ donor3 TT CD4+ donor1 CD4+ donor2 CD4+ donor3 CD4+ donor3

TSS preferences:

- B4GALT1 core promoter
- Primary Astrocytes

~270bp, unprecedented high resolution

33167200

- CD14+ monocytes
- CD4+ T-cells

223,428 in human and 162,264 in mouse of reference TSS

Forrest et al. Nature 507, 462 (2014)

Regulatory network analysis. Why using CAGE?



MOGRIFY: direct cellular reprogramming





Rackham et al. A predictive computational framework for direct reprogramming between human cell types, Nature Genetics, 2016

https://mogrify.co.uk/



More surprises

Bidirectional transcription identifies cell specific enhancers



Systematic in vivo characterization of active enhancers across the human body

- identified 65,423 and 44,459 enhancers in human and mouse.
- 60% are over-represented in one cell/tissue group.
- Human GWAS SNPs map often on enhancers and promoters (less frequently on coding exons).
- Promoter and enhancer usage and QTL analysis. Garieri et al. Nat Commun. (2017)



Andersson et al. Nature 507, 455 (2014)

Disease-associated SNPs are enriched in enhancers

Soluble levels of adhesion molecules Soluble ICAM-1 Hemoglobin Red blood cell traits Primary sclerosing cholangitis Hematological parameters Other erythrocyte phenotypes Fibrinogen Creatinine levels Marfarin maintenance dose Age-related macular degeneration Chronic kidnev disease

GWAS-SNP over-representation in different genomic regions

GWAS SNP sets

Mapping the genome regions that regulate gene activity in diseases

Transcriptome for ALL human cells

 Promoters, enhancers, IncRNAs as FANTOM5 but in "single cells": from cell classification to precision genomics



BULK RNA SEQUENCING

Sequencing a mixture of seemingly identical cells fails to capture the diversity of the immune cells surrounding a tumour.

SINGLE-CELL GENOMICS

Using single-cell genomics, biologists can capture the molecular signature of all immune cells found in and around the tumour.

Giladi et al. Nature (2017)

Taking advantage of single cell work- single cell CAGE

CAGE approach to map regulatory elements and map Predispositions to Diseases



Slide, Chung Chau Hon

The Focus : Cis-Regulatory Elements (CRE) at SINGLE CELL



Slide, Chung Chau Hon

We can now profile transcription and regulatory elements in single cells





Enhancers : Both random and oligo-dT priming detects eRNA in single cells





single base resolution in single cells

From Kouno T., Moody J., Kwon AT, et al. *Nature Communications* 2019

Technology: to identify regulatory elements in genome



Yes CAGE works at single cell, can detect expression, promoters, enhancers, lncRNAs...

Slide, Chung Chau Hon and Jonathan Moody

GWAS interpretation : Linking GWAS variants to candidate genes

We infer how genome sequence variants influence gene expression, in health and diseases, in all human cells





PTGER4 locus

chr5:40,349,192-40,804,356 (455.2kb)

Chung Chau Hon

Comprehensively linking gene expression to human genetics

- > eQTL with much higher resolution at single cell level
- > unbiased cell-type specific expression controlled by genomic elements
- easier to understand if a given genome variation is responsible for the onset/progression of a disease
- eQTL needs ~100 individuals for each cell type to be profiled


Human Cell Atlas (HCA)

Comprehensive reference maps of all human cells at single cell level



A periodic table of our cells



CODEX	GeoMX DSP	tCy-CIF	ST	DNA microsc.	GeoMX DSP	MERFISH	scRNA Seq	ATAC Seq
MIBI	Multiplex IF		Slide Seq	HDST	STAR MAP	Seq FISH	snRNA Seq	DroNc Seq

Graphics credit: Ania Hupalowska

The 7th HCA General Meeting in Japan



Coordination of HCA in Japan and in Asia (HCA executive office at RIKEN)

Back to the non-coding RNAs (Is this a good name?)

CAGE to recount number of IncRNAs

Recounting IncRNAs









Hon et al., Nature 543, 199 (2017

Function:

Do we need experimental evidence?

FANTOM6: Functional IncRNA catalogue



Measuring transcriptional phenotypes

Molecular phenotype induced by IncRNA KD



Characterization of the "new continent" of IncRNA:

- IncRNA functional
- Regulatory role among others
- Many genes with natural antisense RNAs

Ramilowski et al. Genome Res. 30, 1060 (2020)

Jay W. Shin, Michiel de Hoon, Jordan Ramilowski

Example: KD of ZNF213-AS1 impacts cell growth and migration

Wound healing assay for ASOs targeting ZNF213-AS1 35-73% impairment of wound-closure in fibroblasts lacked ZNF213-AS1 Negative Control (NC_A) ASO1 ASO2 ASO3 Image: Ima

Selected enriched biological pathways



45

After thousands of experiments: many IncRNAs have function



Slide courtesy of Jay Shin and Chi Wai Yip

How to collect information about IncRNAs,

faster?

Can we collect how and where IncRNAs interact, their "interactome"?

RNA-DNA interactions affect epigenome and gene regulation



37% of IncRNAs chromatin-bound

What is the role of RNA-chromatin interaction *in cis* and *in trans*?

- Activate genes? Promoter? Enhancer?
- Repress genes? Insulator?

RADICL-seq captures RNA-DNA interactions in intact crosslinked nuclei



mESCs, crosslinked with Formaldehyde.

Other cell types that have been successfully tested with RADICL seq: miPSCs, mMEF, mOPC. Typical seq. depth: 1, ane HiSeq2500/each replica.



Bonetti et al. Nature Communications 11, 1018 (2020)

Interactome analysis: mostly introns in cis; IncRNAs interact more often in trans



RNA sequence and structure unknown, need full-length RNA seq

Different cell types with different interactions patterns of IncRNAs in trans



Slide courtesy of Alessandro Bonetti

Bonetti et al. Nature Communications 11, 1018 (2020)

Cell models and FANTOM6 RADICL libraries (~15 cell types)



In trans interactions increase along differentiation process; IncRNAs and intronic RNAs have different targets



Signals sent through RNAs from gene to other genesand regulatory elements?

A new type of crosstalk: RNA - genome in the nuclei

Further characterization of functions: - structural? - (epi)genome control?



More unexpected function of IncRNAs and repetitive elements

- COI disclosure
- I am co-founder of Transine Therapeutics, a UK company that aims to develop therapeutics based on the SINEUP technology discussed in the presentation. <u>https://www.transinetx.com/</u>

SINEUPs: an important example of antisense RNA function

AS Uch-I1 regulates endogenous UCH-L1 protein expression Uch-I1 AS Uch-I1 mRNA (original non-coding RNA) pcDNA3-AS Uch-I1 (cDNA in plasmid) **qRT-PCR** 1.3 60 0.8 AS Uch-I1 Uch-I1 0.6 30 0.4 20 0.2 10 pcDNA3-AS Uch-I1 Empty vector pcDNA3-AS Uch-I1 Empty vector Western blotting



No changes at RNA level

Protein level: dramatically enhanced!!

Carrieri et al, Nature 491, 454 (2012)

SINEUPs: an important example of antisense RNA function



Carrieri et al, Nature 491, 454 (2012)

SINEUPs

Antisense IncRNAs for therapy

Using SINEUPs (antisense IncRNAs)



Customizable design for any proteins

- Up-regulates protein synthesis 2-5 folds (physiological range)
- Acts on endogenous mRNA and exogenous targets

SINEUPs: in vivo model of human diseases in Medaka

Microphtalmia with Linear Skin Lesions (MLS)



SINEUPs Therapeutics – haploinsufficiencies & others

At least 300 haploinsufficiency genes known (Dang *et al.*, European Journal of Human Genetics, DOI: 10.1038/ejhg.2008.111, 2008)



Regulating genome activity

- Many IncRNAs have regulatory + structural role, sometimes other functions.
 - Large scale ahead
- We always need novel technologies to answer new questions in gene regulation
 - Including computational approaches to measure large networks
- Common diseases are caused by small unbalances of gene regulation for long time. IncRNA are the new frontier for drug development.

Protein coding targeting, well known



Future opportunities in gene regulation

https://humantechnopole.it/en/

R

RIKEN



Piero Carninci

Genomics Research Center, Human Technopole, Milan, Italy RIKEN IMS, Yokohama

- Creating bridges between Japanese and Italian science
- New positions and chances to collaborate





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Thank you

The proportion of noncoding DNA broadly increases with developmental complexity



J.S. Mattick *Nature Reviews Genetics* 5, 316-323 (2004) R.J. Taft, M. Pheasant and J.S. Mattick, *Bioessays* 29, 288-299 (2007)

Non-coding RNA expression in mouse brain



Subcellular localization of ncRNAs



Subcellular localisation of long ncRNAs in Purkinje cells Subcellular localization ~ putative function

2498 Research Article The mRNA-like noncoding RNA Gomafu constitutes a novel nuclear domain in a subset of neurons Masamitsu Sone^{1,2,3}, Tetsutaro Hayashi², Hiroshi Tarui², Kiyokazu Agata², Masatoshi Takeichi^{2,3} and Shinichi Nakagawa^{1,2,*} ¹Nakagawa Initiative Research Unit, RIKEN, 2-1 Hirosawa, Wako 351-0198, Japan ²RIKEN Center for Developmental Biology, 2-2-3 Minatojima Minamimachi, Chuo-ku, Kobe 650-0047, Japan ³Department of Cell and Developmental Biology, Graduate School of Biostudies, Kyoto University, Kitashirakawa, Sakyo-ku, Kyoto 606-8502, Japan *Author for correspondence (e-mail: nakagawas@riken.jp) Accepted 21 May 2007 Journal of Cell Science 120, 2498-2506 Published by The Company of Biologists 2007 doi:10.1242/jcs.009357 B' **B**" Merge DNA Gomafu Α B Gomafu/ DNA

Cell-type-specific IncRNAs implicated in GWAS traits



Hon *et al.*, *Nature* 543, 199 (2017)

Many IncRNAs with potential function

Co-expression of IncRNA-mRNA pairs linked by eQTL





Identified 19,175 potentially functional IncRNAs in human

Hon et al., Nature 543, 199 (2017)

Considerations: localization of IncRNAs And functional interactions



Analyzing tCREs : Gene expression + enhancer activity in one assay

SCAFE : Software to define, quantify and link tCREs from 10x 5'data

Make the most of your 10x 5'data.....for **FREE**!





End-to-end solution for 10x 5'data

Slide courtesy of Chung Chau Hon
Progress in HCA



Explore Guides Metadata Pipelines Analysis Tools Contribute APIs

Mapping the Human Body at the Cellular Level

Community generated, multi-omic, open data processed by uniform pipelines





♦ 179 PROJECTS



e.g. Brain: 172 Donors, 466 specimens, 9.9M Estimated cells Blood: 664 Donors, 2.5k specimens, 2.2M Estimated cells Immune: 9 Donors, 69 specimens, 274.2k Estimated cells

As of December 1, 2021, https://data.humancellatlas.org